(-)-Echinobetaine A: Isolation, Structure Elucidation, Synthesis, and SAR Studies on a New Nematocide from a Southern Australian Marine Sponge, Echinodictyum sp.

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A nematocidal agent present in a southern Australian marine sponge of the genus Echinodictyum has been isolated and identified by detailed spectroscopic analysis and total synthesis as the novel betaine (-)-echinobetaine A (6). Preliminary SAR investigations have been undertaken.

During our investigations into new agrochemical agents from Australian marine organisms we examined a sponge specimen, Echinodictyum sp., collected by beam trawl at a depth of 85 m during a 1995 scientific expedition to the Great Australian Bight, Australia. The EtOH extract of this *Echinodictyum* sp. displayed growth inhibitory properties against the bacteria Serratia marcescens, Micrococcus luteus, and Staphylococcus aureus, but more importantly it displayed excellent in vitro antiparasitic activity against the endo parasite Haemonchus contortus. The latter parasite inflicts serious economic damage to the agricultural sector worldwide through its impact on the health and productivity of livestock such as sheep. While agrochemicals exist to combat such parasites, the increasing incidence of resistance requires that the search for new and improved antiparasitics be both vigorous and ongoing. Experience has shown us that new antiparasitic lead compounds can be discovered through the bioassay-directed exploration of marine biodiversity,¹⁻⁹ with *Echinodictyum* being a promising target for just such an investigation.

Before embarking on this study we reviewed the existing chemical literature on the genus Echinodictyum. The first published account (in 1983)¹⁰ reported the bioassaydirected isolation and subsequent identification of the previously known synthetic compound 4-amino-5-bromopyrrolo[2,3-d]pyrimidine (1), as a potential bronchodilator. To the best of our knowledge the only other report of novel metabolites from an Echinodictyum sp. was an early account by us in 1999.7 In that report we described a series of novel antibacterial agents, echinosulfonic acids A-C (2-4) and echinosulfone (5). At that time our primary interest in Echinodictyum focused on the in vitro antiparasitic activity displayed by the EtOH extracts of at least two specimens in our collection, both from the Great Australian Bight. Although the echinosulfonic acids 2-4 and echinosulfone (5) were identified as natural antibacterial agents, these compounds were not responsible for the antiparasitic properties displayed by the *Echinodictyum* extract. After our earlier report, we persisted in our investigations to the point where we now report the isolation, characterization,

structure elucidation, and synthesis of a nematocidal agent, (-)-echinobetaine A (6), from this same *Echinodictyum* sp.



Results and Discussion

The EtOH extract of the *Echinodictyum* specimen was decanted, concentrated in vacuo, and triturated with CH₂-Cl₂, after which the residue was partitioned between n-BuOH and H₂O. Whereas the n-BuOH solubles displayed antibacterial activity and ultimately led to the isolation and identification of the echinosulfonic acids A-C (2-4) and echinosulfone (5),⁷ the crude H₂O solubles displayed significant antiparasitic activity against H. contortus.

The H₂O solubles were further fractionated by elution through Sephadex G-10 (H_2O) followed by C_{18} HPLC (0.1%)TFA in H₂O) to yield the nematocide (-)-echinobetaine A (6) (*H. contortus*, LD₉₉ 83 µg/mL).

High-resolution ESI(+)MS analysis of 6 revealed a highest mass ion at m/z 176 measuring for C₈H₁₈NO₃ (Δ -0.4 mmu). Examination of the NMR (D₂O, 400 MHz) data for **6** revealed resonances consistent with an OMe (¹H: δ 3.19 (s); ¹³C: 58.6 ppm), an NMe₃ (¹H: δ 2.97 (s); ¹³C: 53.3 ppm), and two deshielded diastereotopic methylenes (¹H: δ 3.31 (dd, J = 13.6, 1.6 Hz) and 3.78 (dd, J = 13.6, 8.0 Hz); ¹³C: 65.1 ppm; as well as ¹H: δ 3.56 (m); ¹³C: 71.8 ppm) flanking a deshielded methine (¹H: δ 3.15 (m); ¹³C: 41.6 ppm), accounting for all but CO_2H of the proposed molecular formula. Furthermore, the appearance of a ¹³C NMR resonance at 175.1 ppm was consistent with the remaining functional unit in 6 being a carboxylic acid. Given the deshielded character of the methylene carbons, and the co-occurrence of quaternary ammonium and carboxylic acid functionalities, the most probable structure for (-)-echinobetaine A (6) was that shown. Analysis of the

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Table 1. NMR (D₂O, 400 MHz) Data for (–)-Echinobetaine A (6)

	$^{13}\mathrm{C}\;\delta$	${}^{1}\mathrm{H}\delta(\mathrm{m},J\mathrm{Hz})$	COSY	$\substack{gHMBC\\(^1H-^{13}C)}$
1	175.1			
2	41.6	3.15 (m)	$3-H_a$, $3-H_b$, $4-H_2$	
$3-H_a$	65.1	3.78 (dd. J = 13.6,	H-2, $3-H_{b}$	NMe ₃ , C-1
		8.0 Hz)		
$3-H_b$		3.31 (dd, J = 13.6,	2-H, 3-H _a	NMe ₃ , C-1
		1.6 Hz)		
NMe_3	53.3	2.97(s)		$C-3$, NMe_3
4	71.8	3.56 (m)	2-H	C-3, OMe
OMe	58.6	3.19 (s)		C-4







2D NMR COSY and gHMBC data for **6** (see Table 1) supported this assignment; however, to provide unambiguous evidence, we undertook the total synthesis of (\pm) -echinobetaine A (16) as outlined in Scheme 1.

Preparation of the benzoate intermediate 13 proceeded in six steps following the method of Bosies et al. (see Scheme 1).¹¹ Quantitative conversion of the readily available diester 7 to the acetal 8 was followed by quantitative conversion to the ester 9. Decarboxylation of 9 yielded 10 as a mixture of *cis* and *trans* isomers (3:4), while subsequent reduction and methylation returned the alcohol 11 (96%) and methyl ether 12 (91%), respectively. Opening of the acetal in 12 with NBS yielded the key intermediate 13 (95%), which was smoothly converted to the corresponding trimethylammonium 14 (98%). Acid hydrolysis of the benzoate ester moiety in 14 returned the alcohol 15, while unoptimized oxidation of 15 returned (\pm)-echinobetaine A (16) in 25% yield after purification by HPLC. Synthetic (\pm)echinobetaine A (16) was identical with natural (-)- Scheme 2. Synthesis of the (\pm) -2-Methoxy- γ -aminobutyric Acid Betaine Methyl Ether $(17)^a$



 $^a\,$ (a) Ag_2O, MeI, 99%; (b) NaOH, 100%; (c) PhCH_2Br, DMF, 30%; (d) i PTsOH, ii NMe_3, 56%.

echinobetaine A (6) by NMR, ESI(+)MS, and IR and by coelution on HPLC.

Of particular note, the nematocidal properties for (\pm) echinobetaine A (**16**) were an order of magnitude weaker than those for the natural product (–)-echinobetaine A (**6**) (LD₉₉ 83 µg/mL), highlighting the significance of stereochemistry in the echinobetaine A pharmacophore. Ongoing studies are directed at assignment of absolute stereochemistry to (–)-echinobetaine A (**6**), as well as chiral resolution of (\pm)-echinobetaine A (**16**), to yield authentic synthetic samples of both enantiomers to support future SAR investigations.

In an attempt to further define the nematocidal pharmacophore revealed by echinobetaine $A(\mathbf{6})$ we prepared the isomer 17 in four steps from (\pm) - α -hydroxybutyrolactone (18) (see Scheme 2). In this sequence commercially available 18 was converted to the methyl ether 19, which was subsequently ring opened to the hydroxycarboxylate salt 20. The salt 20 was then converted to the benzyl ester 21, which was in turn transformed into the target product, (\pm) -2-methoxy- γ -aminobutyric acid betaine methyl ether (17). The isomeric synthetic analogue 17 displayed only very modest nematocidal activity (LD₉₉ 438 μ g/mL), whereas all intermediates prepared during the synthesis of 16 and 17 were inactive. From this bioassay data it would appear that the (-)-echinobetaine A (6) nematocidal pharamacophore does not extend to isomeric methoxy betaines in general and does appear to be stereo-dependent.

(-)-Echinobetaine A (**6**) represents the first example of a new class of antiparasitic natural products and illustrates the all too often unrealized and untapped potential of small molecular weight water-soluble marine metabolites. Given the promising antiparasitic activity displayed by the original *Echinodictyum* extract, we are confident that as yet unidentified water-soluble metabolites with even greater antiparasitic activity await discovery. Our future efforts will extend synthetic studies to further explore and define the (-)-echinobetaine A (**6**) pharmacophore, as well as support ongoing exploration of *Echinodictyum* extracts to detect, isolate, and identify new and hopefully more potent antiparasitics.

Experimental Section

General Experimental Procedures. As previously described.¹²

Animal Material. An *Echinodictyum* sp. (Museum of Victoria Registry Number MVF88741) of marine sponge was collected during a scientific expedition to the Great Australian Bight aboard the *RV Franklin* in July 1995, as previously described.⁷

Extraction and Isolation. The EtOH extract of the *Echinodictyum* specimen was decanted and concentrated in vacuo to yield a bright yellow-orange solid (4.08 g), which was

(-)-Echinobetaine A (6): $[\alpha]^{22}_{D} - 49^{\circ}$ (c 0.59, MeOH); IR (neat) ν_{max} 1683 cm⁻¹; ¹H NMR (400 MHz, D₂O) and ¹³C NMR (100 MHz, D₂O), see Table 1; HRESI(+) MS *m/z* 176.1283 (calcd for C₈H₁₈NO₃, 176.1287).

Diethyl 6-Phenyl-[1,5]-dioxane-3,3-dicarboxylic Acid Diester (8). A solution of diethyl bis-hydroxymethylmalonate (7) (22.0 g, 0.1 mol), PhCHO (10.6 g, 0.1 mol), and *p*-TsOH (0.2 g) was refluxed in toluene (200 mL) for 24 h using a Dean Stark trap, after which the toluene was removed in vacuo and the residue purified by distillation to afford the acetal **8** (7.64 g, 25%): bp_{1mmHg} 160–170 °C; IR (Nujol) ν_{max} 1741 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 7.43–7.31 (m, Ph), 5.46 (s, 6-H), 4.83 (d, J = 11.5 Hz, 2- and 4-H_{2a}), 4.31 (q, J = 7.1 Hz, CO₂CH₂CH₃), 1.29 (t, J = 7.1 Hz, CO₂CH₂CH₃), 1.23 (t, J =7.1 Hz, CO₂CH₂CH₃); ¹³C NMR (75 MHz, CDCl₃) δ 167.6 and 166.7 (2 × CO₂CH₂CH₃), 137.3, 129.0, 128.1, and 126.0 (Ph), 101.6 (C-6), 69.4 (C-4 and C-2), 61.9 (CO₂CH₂CH₃), 53.1 (C-3), 13.9 (CO₂CH₂CH₃); ESI(+)MS *m*/z 331 (M + Na).

Ethyl 6-Phenyl-[1,5]-dioxane-3,3-dicarboxylic Acid Monoester (9). An EtOH solution of the acetal 8 (6.58 g, 0.021 mol) and KOH (1.71 g, 0.03 mol) was stirred at RT for 4 h, after which the EtOH was evaporated in vacuo and the residue treated with 1 M HCl (40 mL) to return the monoester 9 as a white solid (5.50 g, 95%): IR (KBr) ν_{max} 3280–2544 (br), 1741, 1716 cm⁻¹; ¹H NMR (400 MHz, CDCl₃) δ 7.45–7.35 (m, Ph), 6.03 (s, CO₂H), 5.50 (s, 6-H), 4.88 (d, J = 11.6 Hz, 2- and 4-H_{2a}), 4.35 (q, J = 7.2 Hz, CO₂CH₂CH₃), 4.18 (d, J = 11.6 Hz, 2- and 4-H_{2b}), 1.34 (t, J = 7.2 Hz, CO₂CH₂CH₃); ¹³C NMR (75 MHz, CDCl₃) δ 171.9 (CO₂H), 167.5 (CO₂CH₂CH₃), 137.1, 129.3, 128.3, and 126.1 (Ph),101.8 (C-6), 69.3 (C-4 and C-2), 62.5 (CO₂CH₂CH₃), 53.2 (C-3), 14.0 (CO₂CH₂CH₃); ESI(+)MS *m*/z 303 (M + Na).

Ethyl 6-Phenyl-[1,5]-dioxane-3-carboxylic Acid Ester (10). An anhydrous pyridine solution (10 mL) of the ester 9 (5.0 g, 0.018 mol) and piperidine $(30 \,\mu\text{L})$ was refluxed for 5 h, after which the solution was added dropwise to ice cold HCl_{aq} (37%, 13 mL). The resulting solution was extracted with CH₂- Cl_2 (3 imes 50 mL) and the combined organic layer dried (anhydrous MgSO₄) and concentrated in vacuo to afford the decarboxylated product 10 as a mixture of minor (43%) and major (57%) isomers (3.0 g, 72%): IR (Nujol) ν_{max} 1736 cm⁻¹; *minor isomer* ¹H NMR (300 MHz, CDCl₃) δ 7.49–7.35 (m, Ph), 5.52 (s, 6-H), 4.73 (d, J= 11.0 Hz, 2- and 4-H_2a), 4.28 (q, J=7.0 Hz, $CO_2CH_2CH_3$), 4.11 (dd, J = 11.0, 3.3 Hz, 2- and 4-H_{2b}), 2.41 (br m, 3-H), 1.32 (t, J = 7.0 Hz, $CO_2CH_2CH_3$); major isomer ¹H NMR *δ*7.49–7.35 (m, Ph), 5.44 (s, 6-H), 4.47 (dd, J = 11.9, 4.8 Hz, 2- and 4-H_{2a}), 4.16 (q, J = 7.0 Hz, CO_2CH_2 -CH₃), 4.00 (dd, J = 11.9, 11.9 Hz, 2- and 4-H_{2b}), 3.42 (m, 3-H) 1.23 (t, J = 7.0 Hz, CO₂CH₂CH₃); ¹³C NMR (75 MHz, CDCl₃) δ 171.1 and 169.9 (2 \times CO_2CH_2CH_3), 138.0, 137.7, 129.0, 128.9, 128.2, 128.1, 126.1, and 126.0 (Ph), 101.8 and 101.3 (2 × C-6), 68.0 and 67.1 (2 \times C-4 and C-2), 61.1 and 60.8 (2 \times CO₂CH₂-CH₃), 39.9 (C-3), 14.1 ($2 \times CO_2CH_2CH_3$); ESI(+)MS m/z 259 (M + Na).

(6-Phenyl-[1,5]-dioxan-3-yl)methanol (11). The monoester 10 (3.3 g, 0.014 mol) was treated with LiAlH₄ (2.3 g, 0.061 mol) in refluxing dry Et₂O (20 mL) under a N₂ atmosphere for 3 h. The reaction was then quenched with ice and following the dropwise addition of EtOAc (18 mL), water (35 mL), and 15% NaOH (9 mL) was extracted with Et₂O (3 × 50 mL). After drying (anhydrous MgSO₄) the organic phase was concentrated in vacuo to yield the alcohol 11 (2.5 g, 94%) as minor (42%) and major (58%) isomers: IR (Nujol) ν_{max} 3390–3308 (br) cm⁻¹; minor isomer ¹H NMR (400 MHz, CDCl₃) δ 7.50–7.32 (m, Ph), 5.52 (s, 6-H), 4.25 (d, J = 11.2 Hz, 2- and

4-H_{2a}), 4.15–4.11 (m, 2- and 4-H_{2b}), 4.06 (d, J = 7.6 Hz, CH_2 -OH), 1.69 (br s, 3-H); major isomer ¹H NMR (400 MHz, CDCl₃) δ 7.50–7.32 (m, Ph), 5.43 (s, 6-H), 4.32 (dd, J = 11.6, 4.8 Hz, 2- and 4-H_{2a}), 3.74 (dd, J = 11.6, 11.6 Hz, 2- and 4-H_{2b}), 3.52 (d, J = 6.0 Hz, CH_2 OH), 2.39 (m, 3-H); ¹³C NMR (100 MHz, CDCl₃) δ 138.3, 138.1, 128.9, 128.8, 128.2, 126.0, and 125.9 (Ph), 101.8 and 101.4 (2 × C-6), 69.6 and 67.7 (2 × C-4 and C-2), 61.5 and 60.9 (2 × CH_2 OH), 36.8 and 36.4 (2 × C-3); ESI-(+)MS m/z 217 (M + Na).

3-Methoxymethyl-6-phenyl-[1,5]-dioxane (12). A dry THF solution of the alcohol 11 (2.48 g, 0.013 mol) and NaH (1.53 g, 0.064 mol) was refluxed under a N₂ atmosphere for 1 h, after which MeI (9 mL, 0.11 mol) was added dropwise and the reaction refluxed for a further 2 h. On cooling the reaction mixture was extracted with $Et_2O(3 \times 50 \text{ mL})$, and the organic phase washed with brine $(3 \times 20 \text{ mL})$, dried (anhydrous $MgSO_4$), and concentrated in vacuo to yield the methyl ether $12\ (2.46\ g,\ 91\%)$ as minor (36%) and major (64%) isomers: *minor isomer* ¹H NMR (400 MHz, CDCl₃) δ 7.50-7.31 (m, Ph), 5.50 (s, 6-H), 4.21 (d, J = 10.8 Hz, 2- and 4-H_{2a}), 4.11-4.08 (m, 2- and 4-H_{2b}), 3.76 (d, J = 7.2 Hz, CH₂OMe), 3.41 (s, CH₂OMe), 1.74 (br m, 3-H); major isomer ¹H NMR (400 MHz, $CDCl_3$) δ 7.50–7.32 (m, Ph), 5.42 (s, 6-H), 4.28 (dd, J = 11.8, 4.4 Hz, 2- and 4-H_{2a}), 3.73 (dd, J = 11.8, 11.8 Hz, 2- and 4-H_{2b}), 3.32 (s, CH₂OMe), 3.24 (d, J = 5.6 Hz, CH₂OH), 2.47 (m, 3-H); ¹³C NMR (75 MHz, CDCl₃) δ 138.4, 138.2, 128.5, 127.9, 125.8, and 125.7 (Ph), 101.5 and 101.1 (2 \times C-6), 71.3 and 70.6 (2 \times CH₂OMe), 69.5 and 67.6 (2 \times C-4 and C-2), 58.7 and 58.6 (2 \times CH_2OMe), 34.6 and 34.4 (2 × C-3); ESI(+)MS m/z 231 (M + Na)

3-Bromo-2-(methoxymethyl)-1-propyl Benzoate (13). A mixture of the methyl ether 12 (2.46 g, 0.012 mol), freshly recrystallized N-bromosuccimide (2.2 g, 0.006 mol), and BaCO₃ (0.359 g, 0.002 mol) was refluxed in dry CH₂Cl₂ (30 mL) for 24 h under a N₂ atmosphere. After cooling, the reaction was concentrated in vacuo and the residue triturated with hexane, filtered, and dried in vacuo to yield the bromobenzoate 13 (3.3) g, 95%): IR (neat) $\nu_{\rm max}$ 1724 cm^-
i; ¹H NMR (300 MHz, CDCl_3) δ 8.04 (d, J = 7.4 Hz, 2'-H and 6'-H), 7.58 (t, J = 7.4 Hz, 4'-H), 7.46 (t, J = 7.4 Hz, 3'-H and 5'-H), 4.45 (dd, J = 11.2, 6.4 Hz, 1-H_{2a}), 4.38 (dd, J = 11.2, 6.4 Hz, 1-H_{2b}), 3.61 (d, J = 5.6Hz, 4-H₂), 3.56 (dd, J = 7.8, 5.0 Hz, 3-H_{2a}), 3.51 (dd, J = 7.8, $5.0~{\rm Hz},\,3\text{-}{\rm H_{2b}}),\,3.37~(s,\,OMe),\,2.47~(m,\,2\text{-}{\rm H});\,^{13}\!C~NMR~(75~{\rm MHz},$ CDCl₃) & 166.0 (CO₂Ph), 132.9, 129.8, 129.4, and 128.3 (CO₂Ph), 70.9 (C-4), 63.6 (C-1), 58.9 (OMe), 40.5 (C-2), 31.9 (C-3); ESI(+)MS m/z 309/311 (M + Na).

(±)-3-Trimethylammonium-2-(methoxymethyl)-1-propyl) Benzoate (14). A solution of the bromobenzoate 13 (3.3 g, 0.013 mol) in 23% trimethylamine in dry benzene (200 mL) was stirred for 48 h, after which the resulting precipitate was filtered and washed with Et₂O (50 mL) to yield the insoluble trimethylammonium salt 14 (3.40 g, 98%). Recrystallization from methanol/EtOAc returned a white solid with a melting point of 125–127 °C: IR (Nujol) ν_{max} 1720 cm⁻¹; UV (MeOH) $\bar{\lambda}_{\max}$ (ϵ) 229 (7400); ¹H NMR (400 MHz, CDCl₃) δ 7.98 (d, J =7.8 Hz, 2'-H and 6'-H), 7.56 (t, J = 7.8 Hz, 4'-H), 7.43 (t, J =7.8 Hz, 3'-H and 5'-H), 4.55 (dd, J = 11.8, 5.2 Hz, 1-H_{2a}), 4.34 $(dd, J = 11.8, 7.2 Hz, 1-H_{2b}), 3.86 (dd, J = 13.4, 4.4 Hz, 3-H_{2a}),$ $3.70 \,(dd, J = 13.4, 3.2 \,Hz, 3-H_{2b}), 3.61 \,(dd, J = 9.2, 4.4 \,Hz,$ 4-H_{2a}), 3.51 (s, NMe₃), 3.46 (dd, J = 9.2, 7.2 Hz, 4-H_{2b}), 3.34 (s, OMe), 2.71 (br m, 2-H); 13 C NMR (100 MHz, CDCl₃) δ 166.3 (CO₂Ph), 133.3, 129.5, 129.1, and 128.4 (CO₂Ph), 71.4 (C-4), 65.8 (C-3), 64.3 (C-1), 59.1 (OMe), 53.6 (NMe₃), 34.7 (C-2); HRESI(+)MS m/z 266.1764 (calcd for C₁₅H₂₄NO₃, 266.1756).

(±)-3-Trimethylammonium-2-(methoxymethyl)-1-propanol (15). The trimethylammonium salt 14 (1.65 g, 0.006 mol) was refluxed for 24 h in 1 M HCl (80 mL), then washed with EtOAc, and the aqueous layer was concentrated in vacuo to yield the alcohol 15 (840 mg, 87%). Recrystallization from MeOH/EtOAc returned flake-like crystals with a melting point of 123–124 °C: IR (Nujol) ν_{max} 3275 cm⁻¹; ¹H NMR (400 MHz, D₂O) δ 3.49 (dd, J = 11.6, 5.7 Hz, 3-H_{2a}), 3.42 (dd, J = 11.6, 5.7 Hz, 3-H_{2b}), 3.36 (dd, J = 10.4, 5.2 Hz, 1-H_{2a}), 3.30 (dd, J = 10.4, 6.4 Hz, 1-H_{2b}), 3.19 (d, J = 4.0, Hz, 4-H₂), 3.23 (s, OMe), 3.00 (s, NMe₃), 2.19 (m, 2-H); ¹³C NMR (100 MHz, D₂O) δ 71.5

(C-4), 65.5 (C-3), 60.9 (C-1), 58.4 (OMe), 53.2 (NMe₃), 36.0 (C-2); HRESI(+)MS m/z 162.1489 (calcd for C₈H₂₀NO₂, 162.1494).

(\pm)-Echinobetaine A (16). A 54% AcOH_{aq} solution of the alcohol 15 (0.142 g, 0.97 mmol) and KMnO₄ (0.278 g, 1.8 mmol) was stirred for 3 h at 50 °C, after which the reaction was filtered and concentrated in vacuo. The resultant solid was eluted through a C_{18} solid-phase extraction (SPE) cartridge with H₂O. The solvent was removed in vacuo and the residue subjected to isocratic C_{18} HPLC (2.0 mL/min $H_2O + 0.1\%$ TFA, Zorbax 10 μ m 250 \times 10 mm column) to yield (±)-echinobetaine A (16) (43 mg, 25%): IR, ¹H NMR (400 MHz, D₂O), and ¹³C NMR (100 MHz, D_2O) identical with (-)-echinobetaine A (6); HRESI(+)MS m/z 176.1282 (calcd for C₈H₁₈NO₃, 176.1287).

 (\pm) -2-Methoxy- γ -butyrolactone (19). A solution of the (\pm) -2-hydroxy- γ -butyrolactone (18) (1.87 g, 18.3 mmol), MeI (11 mL, 176 mmol), and Ag₂O (17.8 g, 77 mmol) was refluxed under a N2 atmosphere in CHCl3 (50 mL) for 1 h. The reaction was cooled and filtered, and the clear solution was concentrated in vacuo to yield the methoxy butyrolactone 19 (2.10 g, 99%): ¹H NMR (CDCl₃, 400 MHz) δ 4.40 (m, 4-H_{2a}), 4.23 (m, $4-H_{2b}$), 4.02 (t, J = 7.6 Hz, 2-H), 3.56 (s, OMe), 2.50 (m, $3-H_{2a}$), 2.23 (m, 3-H_{2b}); ¹³C NMR (75 MHz, CDCl₃) δ 174.6 (C-1), 74.6 (C-2), 65.0 (C-4), 57.6 (OMe), 28.9 (C-3).

Sodium (±)-4-Hydroxy-2-methoxybutyrate (20). A solution of the methoxy butyrolactone 19 (2.10 g, 18.1 mmol) in 2 M NaOH (6 mL) and MeOH (6 mL) was stirred for 16 h at RT and the reaction mixture concentrated in vacuo to return the crude sodium salt 20 (2.80 g, 100%), which was used without further purification: ¹H NMR (D₂O, 300 MHz) δ 3.56 (dd, J =8.1, 4.8 Hz, 2-H), 3.48 (t, J = 7.2 Hz, 4-H₂), 3.12 (s, OMe), 1.70 (m, 3-H₂); 13 C NMR (D₂O, 75 MHz) δ 180.3 (C-1), 79.9 (C-2), 58.2 (C-4), 57.0 (OMe), 35.0 (C-3).

Benzyl (\pm) -4-Hydroxy-2-methoxybutanoate (21). The sodium salt 20 (2.80 g, 18.00 mmol), benzyl bromide (2.2 mL, 18.7 mmol), and 18-crown-6-(1,4,7,10,13,16-hexaoxacyclooctadecane) (1.60 g, 6.2 mmol) were stirred at 40 °C in DMF (30 mL) under a N₂ atmosphere for 4 days, after which the reaction mixture was quenched with H₂O (30 mL), extracted with Et₂O, washed with brine, dried over anhydrous Mg₂SO₄, and concentrated in vacuo to yield the benzyl ester 21 (1.20 g, 30%), which was purified by rapid silica filtration eluting with 30% ethyl acetate/petroleum spirit: ¹H NMR (CDCl₃, 400 MHz) δ 7.37 (m, OCH₂Ph), 5.23 (d, J = 12.4 Hz, OCH_{2a}Ph), 5.18 (d, J = 12.4 Hz, OCH_{2b}Ph), 4.03 (dd, J = 7.6, 4.8 Hz, 2-H), 3.76 (t, J = 5.2 Hz, 4-H₂), 3.43 (s, OMe), 2.01 (m, 3-H₂); ¹³C NMR (D₂O, 100 MHz) & 172.2 (C-1), 128.6, 128.5, 128.4, and 128.3 (OCH₂Ph), (79.1 (C-2), 66.7 (OCH₂Ph), 59.7 (C-4), 58.4 (OCH₃), 35.1 (C-3).

(±)-2-Methoxy-γ-aminobutyric Acid Betaine Methyl Ether (17). A solution of the benzyl ester 21 (1.20 g, 5.35 mmol) and *p*-toluenesulfonyl chloride (2.04 g, 10.7 mmol) was stirred in dry pyridine at 5 °C for 3 h. Diethyl ether was added and the organic phase washed sequentially with water, 5% copper sulfate, and water, after which it was dried with anhydrous MgSO₄ and concentrated in vacuo to yield a 4-tosyl intermediate (1.39 g, 68%). The tosyl intermediate was heated to 95 °C with a solution of trimethylamine in benzene in a sealed tube for 16 h, after which the reaction mixture was concentrated in vacuo to a clear, viscous oil (1.60 g, 91%), which was dissolved in 4 M HCl (15 mL) and dioxane (5 mL) and stirred at RT for 16 h. The reaction mixture was concentrated in vacuo to yield a viscous oil, which was purified by C₁₈ SPE (10% stepwise elution from 100% H_2O to 100% MeOH). The 100% H₂O fractions were combined, concentrated in vacuo, and further purified by anion exchange SPE (preconditioned and sample loaded from 10 mL of 0.1 \bar{M} NaCl_aq, elution with $H_2O)$ to yield the trimethylammonium 17 (350 mg, 56%): IR (neat) v_{max} 1736 cm⁻¹; ¹H NMR (D₂O, 400 MHz) δ 3.89 (dd, J = 8.0, 3.6 Hz, 2-H), 3.30 (dt, J = 12.4, 4.8 Hz, 4-H₂), 3.23 (s, OMe), $2.96\,(s,\,NMe_3),\,2.20\,(m,\,3\text{-}H_{2a}),\,2.00\,(m,\,3\text{-}H_{2b});\,^{13}C\,\,NMR\,(D_2O,\,10^{-3})$ 75 MHz) & 174.8 (C-1), 76.8 (C-2), 62.8 (C-4), 58.1 (OMe), 53.1 (NMe₃), 25.5 (C-3); HRESI(+)MS m/z 176.1282 (calcd for C₈H₁₈-NO₃, 176.1287).

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References and Notes

- Capon, R.; Skene, C.; Liu, E. H.-T.; Lacey, E.; Gill, J. H.; Heiland, K.; Friedel, T. J. Nat. Prod. 2004, 67, 1277-1282.
- K.; Friedel, T. J. Nat. Prod. 2004, 67, 1277–1282.
 (2) Capon, R. J.; Skene, C.; Vuong, D.; Lacey, E.; Gill, J. H.; Heiland, K.; Friedel, T. J. Nat. Prod. 2002, 65, 368–370.
 (3) Capon, R. J.; Ford, J.; Lacey, E.; Gill, J. H.; Heiland, K.; Friedel, T. J. Nat. Prod. 2002, 65, 358–363.
 (4) Vuong, D.; Capon, R. J.; Lacey, E.; Gill, J. H.; Heiland, K.; Friedel, T. J. Nat. Prod. 2001, 64, 640–642.
 (5) Capon, R. J.; Shene, C.; Lin, F. H. T.; Lacey, F.; Cill, J. H.; Heiland, K.; Friedel, T. J. Nat. Prod. 2001, 64, 640–642.

- Capon, R. J.; Skene, C.; Liu, E. H.-T.; Lacey, E.; Gill, J. H.; Heiland,
- (a) Capon, R. J., Skene, C., Luc, F. H. 1, Lacy, L., Chi, S. H., Reihard, K., Friedel, T. J. Org. Chem. 2001, 66, 7765–7769.
 (b) Ovenden, S. P. B.; Capon, R. J.; Lacey, E.; Gill, J. H.; Friedel, T.; Wadsworth, D. J. Org. Chem. 1999, 64, 1140–1144.
 (7) Ovenden, S. P. B.; Capon, R. J. J. Nat. Prod. 1999, 62, 1246–1249.
 (8) Chard Chard Charge F. Cill, J. H. Wadsworth, D. Fieldel
- (8) Capon, R. J.; Skene, C.; Lacey, E.; Gill, J. H.; Wadsworth, D.; Friedel, T. J. Nat. Prod. 1999, 62, 1256–1259.
- Capon, R. J.; Barrow, R. A.; Rochfort, S.; Jobling, M.; Skene, C.; Lacey, (9)E.; Gill, J. H.; Friedel, T.; Wadsworth, D. Tetrahedron 1998, 58, 2227-2245.
- (10) Kazlauskas, R.; Murphy, P. T.; Wells, R. J.; Baird-Lambert, J. A.; Jamieson, D. D. Aust. J. Chem. 1983, 36, 165–170.
- (11) Bosies, E.; Herrmann, D. B. J.; Bicker, U.; Gall, R.; Pahlke, W. Lipids
- Jost, 22, 947–951.
 Capon, R. J.; Skene, C.; Stewart, M.; Ford, J.; O'Hair, R. A. J.;
 Williams, L.; Lacey, E.; Gill, J. H.; Heiland, K.; Friedel, T. Org., Biomol. Chem. 2003, 1, 1856–1862. (12)

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